

# A KIR x CD8 targeting bispecific modulator enhances regulatory CD8 T cell functions, and reduces inflammation in models of autoimmune disease

### Orchestrating The Immune System

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#### Introduction

We have characterized a subset of CD8 T cells (CD8 Treg) with immunosuppressive characteristics in inflammatory disease settings. CD8 Treg activation results in their oligoclonal expansion and elimination of pathogenic CD4 T cells. Here we describe a bispecific CD8 Treg Modulator that binds KIR2DL1/2/3 and CD8 to activate autoimmune patient CD8 Tregs resulting in the cytolytic elimination of pathogenic CD4 T cell in vitro, ex vivo, and in vivo.

## **CD8** Treg Modulator is targeting a Novel Network to **Restore Immune Balance in Autoimmune Disease**







CD8 Treg Modulators are designed to restore homeostasis

Approach seeks to increase cytolytic activity of dysfunctional CD8 Treg

#### Methods

We tested target binding of a novel bispecific CD8 Treg Modulator by Octet, and to immune cell subsets in healthy donor PBMC by flow cytometry.

The functional impact of the CD8 Treg Modulator was evaluated in healthy, Celiac, and Crohn's PBMC by CD8 Treg activation, cytotoxicity assays, and supernatant analysis in vitro, with Celiac patient derived biopsies ex vivo, and in acute GVHD studies using healthy donor human PBMC.

#### Simultaneous antigen binding and selective targeting of **CD8 Treg Modulator to dual target expressing CD8 T cells**



Figure 1. Binding of CD8 Treg Modulator to target recombinant proteins as detected by Octet. The CD8 Treg Modulator was captured onto immobilized KIR followed by CD8α binding.



Figure 2. Mixed PBMC from a healthy donor were incubated with the bispecific CD8 Treg Modulator or each of the single arm control molecules. Binding of the antibodies to different cell populations was detected using an anti-human IgG1 Fc secondary antibody. A nonblocking KIR antibody was used to selectively gate on the KIR+ CD8 Tregs within the total PBMC.

# CD8 Treg Modulator preferentially binds to CD8 Tregs inducing their activation with limited NK cell binding and no activation





Figure 3. PBMC from a healthy donor were incubated with KIRxCD8 bispecific CD8 Treg Modulator and binding of the antibodies to different cell populations was detected using an anti-human IgG1 Fc secondary antibody. Healthy donor CD8 Tregs were incubated with the KIRxCD8 bispecific CD8 Treg Modulator for 3 days and then CD8 Treg activation was determined by gating on CD8 Treg with non-blocking KIR2D and CD69 antibodies. KIRxCD8 bispecific CD8 Treg Modulator binding to NK cells was evaluated over a range of doses, and NK cell activation measured following incubation with KIRxCD8 bispecific CD8 Treg Modulator or KIR single arm control antibody.

#### CD8 Treg Modulator enhances the elimination of Ag-responsive CD4s in Celiac and Crohn's PBMC

Celiac PBMC



**INCREASED GRANZYME B** CD8 Treg Modulator (nM

CD8 Treg Modulator pliadin activated untreated con Time (Hrs)



# Crohn's PBMC



Figure 4. CD8 Tregs detected in either healthy or Celiac PBMC. Increase in Granzyme B content in Celiac donor CD8 Tregs following-co-culture with gliadin-stimulated CD4 T cells and 80nM KIRxCD8 bispecific CD8 Treg Modulator for 18 hours. CD8 Treg elimination of celiac-responsive CD4 T cells as detected by a decrease in GFP signal over time by Incucyte. Elimination of gliadin-responsive Celiac donor CD4 T cells as detected by increase in Annexin V+ CD4 T cells at 48hrs following addition of KIRxCD8 bispecific CD8 Treg Modulator and decrease in CD25+ CD4s on Day 5. Increase in Granzyme B content in Crohn's donor CD8 Tregs following flagellin-specific stimulation and treatment with KIRxCD8 bispecific CD8 Treg Modulator. Flagellin-specific activation of Crohn's donor CD4 T cells was reduced by treatment with KIRxCD8 bispecific CD8 Treg Modulator as detected by reduction in CD25<sup>+</sup> CD4 T cells, CXCR3<sup>+</sup> PD-1<sup>+</sup> CD4 T cells, and IL-2.

# CD8 Treg Modulator treatment reduces epithelial cell death in Celiac duodenal biopsy organoid cultures following gliadin-specific stimulation





**INCREASED CD8 TREG** PREVALENCE

Figure 5. Image of an organoid generated from a duodenal biopsy from a Celiac patient that was cultured for ten days in a collagen matrix in the presence of essential growth factors. Organoid were treated, harvested following culture, and processed for flow cytometry. Gliadin peptide stimulation activated CD4 T cells and increased epithelial cell death that was reduced with the addition of CD8 Treg Modulator. CD8 Treg Modulator treatment also expanded CD8 Tregs within the organoid culture.

# CD8 Treg Modulator activates and expands CD8 Tregs in Human PBMC-NSG<sup>TM</sup> GvHD Model









human PBMC followed by weekly doses of CD8 Treg Modulator or Saline control. Peripheral blood was collected at day 14 and CD8 Treg Modulator bound to CD8 Tregs was detected with an anti-human IgG1 Fc secondary antibody. CD8 Treg Modulator treated CD8 Tregs had increased Granzyme B content and a dosedependent increase in the activation marker

# **CD8** Treg modulator ameliorates CD4 mediated pathology in Human PBMC-NSG<sup>TM</sup> GvHD Model





Figure 7. NSG Mice injected with human PBMC followed by weekly doses of CD8 Treg Modulator. Activated CD25+ CD4 T cells and the proinflammatory cytokines GMCSF, IFN<sub>γ</sub>, and TNF $\alpha$  were decreased with increasing dose of CD8 Treg Modulator with a corresponding increase in Annexin V+ CD4 T cells.

## **Dose-dependent concentration of CD8 Treg Modulator** detected in humanized mice PBMC-NSG<sup>TM</sup> GvHD Model

#### SERUM CONCENTRATIONS OF CD8 TREG MODULATOR



Figure 8. Serum was collected at different time points following intravenous injection of different doses of KIRxCD8 bispecific CD8 Treg Modulator. Serum concentration was detected by KIR capture ELISA. n=14 mice at SD0, 2h; n=8 mice at 336h ; up to n=8 mice for terminal time points (ranging from 768 hr to 1008 hr), Interim terminal on 120 hr, 264 hr, n=3/time point Terminal analysis for 0.02 mg/kg group not done (n.d.)

#### \*Please see Mozart Poster #P970: "Pre-clinical pharmacologic and tolerability characterization of a novel KIRxCD8 targeting bispecific CD8 Treg modulator"

- donor derived PBMC.
- CD8 Treg Modulator increases CD8 Treg prevalence and NSG mouse model of acute GVHD.
- diseases.

Acknowledgements

culturing methods, and Jackson laboratories for performing GVHD studies. Contact: Follow up questions can be directed to Kristine Swiderek, Chief Scientific Officer, Mozart Therapeutics kswiderek@mozart-tx.com or by visiting the website at <a href="https://www.mozart-tx.com/">https://www.mozart-tx.com/</a>

References: Li et al. KIR + CD8 + T cells suppress pathogenic T cells and are active in autoimmune diseases and COVID-19. Science. 2022 DOI: 10.1126/science.abi9591



Conclusions

• A novel KIR x CD8 targeting bispecific CD8 Treg Modulator selectively binds and activates CD8 Tregs to restore cytolytic function towards pathogenic CD4 T cells in Celiac and Crohn's

• In ex vivo Celiac duodenal organoid cultures, treatment with the CD8 Treg Modulator increases CD8 Treg prevalence and reduces epithelial cell death induced by gluten-reactive CD4s.

activation while reducing CD4 pathology in a human PBMC

• We postulate that the CD8 Treg Modulator can reduce autoreactive CD4 T cell prevalence to reestablish immune balance and may represent a selective, broadly applicable and novel therapeutic approach for the treatment of autoimmune

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